Opal[™] 4-Color and 7-Color Automation IHC Kits

For Leica Biosystems BOND RX System Software version 4.0

User Manual



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Preface

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Email: global.techsupport@perkinelmer.com

Internet: www.perkinelmer.com

For questions about BOND RX operation, please contact Leica Biosystem Technical support at **844-534-2262**.

Before you call, have the following information available for the technical representative:

- Reagent Lot numbers
- BOND RX Software version
- If applicable, the *error number* shown in the software or in the log file.

Obtaining MSDSs

Material Safety Data Sheets for the reagents in the Opal 4-Color and 7-Color Automation Kits are available on the PerkinElmer website (www.perkinelmer.com). MSDSs for chemicals from other manufacturers should be obtained from the chemical manufacturer.

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NOTE



Changes or modifications to the protocol not expressly approved by PerkinElmer could affect the results of the protocol.

Table of Symbols

Table 1 contains symbols that identify particularly important information and alert you to the presence of hazards. These symbols may appear in this manual and/or on the product it describes.

Table 1. Important Symbols

Symbol Symbole	Description Description
1	DANGER: An imminently hazardous situation, which, if not avoided, will result in death or serious injury. DANGER: Situation présentant un danger imminent qui, s'il n'est pas éliminé, peut entraîner des blessures graves, voire la mort.
<u> </u>	WARNING: Caution, risk of danger. Refer to the User's documentation. AVERTISSEMENT: Attention, danger potentiel. Se reporter à la documentation de l'utilisateur.
!	NOTE: A cautionary statement; an operating tip or maintenance suggestion; may result in instrument damage if not followed. REMARQUE: Énoncé indiquant une précaution à prendre, un conseil de fonctionnement ou une suggestion d'entretien; son non-respect peut provoquer des dommages à l'instrument.
	Risk of exposure to biohazards. Risque d'exposition aux risques biologiques.
	Crush hazard. Risk of body parts, hair, jewelry, or clothing getting caught in a moving part. Danger d'écrasement. Faire attention que les parties corporelles, les cheveux, les bijoux ou les vêtements ne soient pas pris dans une pièce mobile.
	Risk of eye injury; wear safety glasses. Risque de lésion oculaire; porter des lunettes de sécurité.
	Risk of poison. Risque d'empoisonnement.
A	Hazardous fumes. Émanations dangereuses.

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Introduction

Product Description

Opal™ is a method for multiplex fluorescent immunohistochemistry in formalin-fixed, paraffin-embedded (FFPE) tissue. It allows use of standard unlabeled primary antibodies, including multiple antibodies raised in the same species. The basic approach was inspired by a protocol published by Zsuzsanna Tóth and Éva Mezey. The current method involves detection with Opal reactive fluorophores that covalently label the epitope. After labeling is complete, antibodies are removed in a manner that does not disrupt the Opal fluorescence signal. This allows the next target to be detected without fear of antibody cross reactivity. Opal enables development of multiplexed assays with balanced, quantitative signal for rare and abundant targets.

PerkinElmer's Opal 4-Color and 7-Color Automation IHC Kits are for Research Use Only. They are not for Use in Diagnostic Procedures.

Leica Biosystems BOND RX is for Research Use Only. It is not for Use in Diagnostic Procedures.

Opal 4-Color and 7-Color Automation IHC Kits Reagents and Storage

Opal 4-Color or 7-Color Automation IHC Kit contains enough reagents to run 50 standard Opal 4-Color or 7-Color Assays on Leica Biosystems BOND RX System at 1:150 recommended dilution rate for all Opal fluorophores.

The reagents have a shelf life of 12 months from the shipment date when stored using indicated temperatures in Table 1.

Opal 4-Color Automation IHC Kit Quantity Storage Reagent 4°C Antibody Diluent/Block 100 mL 50 mL 4°C Opal Polymer HRP Ms+Rb dry material; 1 vial -20 °C Opal 520 Reagent dry material; -20 °C Opal 570 Reagent 1 vial drv material: 1 vial -20 °C Opal 690 Reagent 4°C Spectral DAPI 1 mL 4°C 1X Plus Amplification Diluent 50 mL Room temperature DMSO (Anhydrous Grade) 500 µL (20-25 °C)

Table 1a. Opal 4-Color Automation IHC Kit

Table 1b. Opal 7-Color Automation IHC Kit

Opal 7-Color Automation IHC Kit				
Reagent	Storage			
Antibody Diluent/Block	100 mL	4 °C		
Opal Polymer HRP Ms+Rb	50 mL; 2 bottles	4 °C		
Opal 520 Reagent	dry material; 1 vial	-20 °C		
Opal 540 Reagent	dry material; 1 vial	-20 °C		
Opal 570 Reagent	dry material; 1 vial	-20 °C		
Opal 620 Reagent	dry material; 1 vial	-20 °C		
Opal 650 Reagent	dry material; 1 vial	-20 °C		
Opal 690 Reagent	dry material; 1 vial	-20 °C		
Spectral DAPI	1 mL	4 °C		
1X Plus Amplification Diluent	50 mL; 2 bottles	4 °C		
DMSO (Anhydrous Grade)	500 μL	Room temperature (20–25 °C)		

BOND RX Materials Required

The Opal 4-Color and 7-Color Automation IHC Assay require the following materials and equipment that are available only from Leica Biosystems and are listed in Table 2.

Table 2. BOND RX Materials Required

Component	Cat. No.	Storage
BOND Titration Kit	OPT9049	Room temperature (20–25 °C)
BOND Open Containers 30 mL	OP309700	Room temperature (20–25 °C)
BOND Open Containers 7 mL	OP79193	Room temperature (20–25 °C)
BOND Research Detection System 2	DS9777	Room temperature (20–25 °C)
BOND Universal Covertiles 160 pack	S21.4611	Room temperature (20–25 °C)
BOND Slide Tray	S21.0304	Room temperature (20–25 °C)
BOND Reagent Tray	S21.1003	Room temperature (20–25 °C)

BOND Slide Labels and Printer Ribbon	S21.4564	Room temperature (20–25 °C)
BOND Apex Adhesive Slide	3800040	Room temperature (20–25 °C)
BOND Dewax Solution – 1L (RTU)	AR9222	2–26 °C
BOND Epitope Retrieval Solution 1-1L (RTU)	AR9961	2–8 °C
BOND Epitope Retrieval Solution 2-1L (RTU)	AR9640	2–8 °C
BOND Wash Solution 10X Concentrate – 1L	AR9590	2–8 °C
BOND Aspirating Probe Cleaning System	CS9100	2–8 °C

Equipment

Component	Cat. No.
Leica Biosystems' BOND RX System - automated slide stainer	
PerkinElmer's Vectra [®] Polaris [™] , Vectra [®] 3 or Mantra [™] Quantitative Pathology Workstation for imaging	

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User-Supplied Materials Required

The materials below are required to run the assay and prepare the slides.

Table 3. User-Supplied Materials Required

Description	Supplier	Cat. No.
200 Proof ethanol (EtOH)	MLS	_
Xylene	MLS	_
10% neutral-buffered formalin (NBF)	MLS	_
Paraffin wax	MLS	_
1X Tris-Buffered Saline (TBS)	MLS	_
Microtome	MLS	_
Drying oven, capable of holding temperature at 65 +/- 1 °C (optional)	MLS	_
Slide Rack 30 - Plastic (optional)	Leica	14047533750
Water bath or incubator, capable of holding temperature at 40 +/- 1 °C	MLS	_
ProLong® Diamond Antifade Mountant	MLS	P36961
Slide Rack	MLS	_
Staining Dish	MLS	_
Cover Glass No. 1.5	MLS	_
Deionized Water	MLS	_
Fume hood	MLS	_

^{*} MLS = Major Laboratory Supplier in North America. For other regions, please check with local vendors.

Before You Begin

Before running either the Opal 4-Color or 7-Color Automation IHC Kit on the Leica Biosystems BOND RX:

- Read and understand the Leica Biosystems BOND RX Instructions for
- Read and understand this document.

Important Guidelines

- Always use properly prepared tissue samples. See Prepare Tissue Samples on page 11 for tissue sample preparation.
- Recommend running the BOND Aspirating Probe Cleaning System more often than conventional IHC.
- Always run control slides with sample slides to verify proper staining.
- Do not use consumables or reagents other than those specified.
- Changing the protocol may affect the results of the run.
- Do not let the tissue samples dry out at any time after BOND RX run is finished.
- Use good laboratory practices and follow all safety procedures. See Safety on page 33 for more information.

Prepare Tissue Samples

This section describes how to prepare and pretreat formalin-fixed, paraffinembedded (FFPE) tissue sections for use with the Opal 4-Color or 7-Color Automation IHC Kit on the BOND RX.

NOTE



Using other preparation or pretreatment procedures may affect the results of the protocol.

Prepare the FFPE Sections

Materials Required

- 10% neutral buffered formalin (NBF)
- 1x TBS
- Paraffin wax
- 200 Proof EtOH
- **Xylene**
- Microtome
- Water bath
- **BOND APEX Adhesive slides**
- Drying oven

Fix the Tissue Sample

Immediately after dissection, fix tissue in 10% NBF for 16-32 hours at ROOM TEMPERATURE (RT). Fixation time will vary depending on tissue type and size.

WARNING



Biological Risk. Handle all samples using appropriate safety procedures.

NOTE



Fixation for less than 16 hours or more than 32 hours will impair the results of the Opal 4-Color or 7-Color Automation IHC Assay.

Dehydrate, Embed, and Cut the Tissue Sample

WARNING



Do not use reagents or consumables that are past their expiration date.

- 1 Wash the sample with 1x TBS.
- 2 Dehydrate the sample using a standard ethanol series, followed by xylene.
- 3 Embed the sample in paraffin using standard procedures. Note that paraffin-embedded samples can be stored at RT.
- 4 Trim paraffin blocks and then cut the embedded tissue into $5 \pm 1 \mu m$ sections using a microtome.
- Place the paraffin ribbon in a 35-45 °C water bath and mount sections on BOND Apex Adhesive slides or Superfrost® Plus Micro Slide.
- 6 Load slides onto the vertical 24-Slide Rack and air dry overnight.
- If desired, the sectioned tissue can be stored with desiccants at RT before continuing to run the Opal 4-Color or 7-Color Automation IHC Kit.
- Bake slides at 65 °C for 1 hour in drying oven before staining.

NOTE



Tip the slide rack to one side and place paper towels underneath to soak up the wax while drying.

Remove and let cool.

Opal Reagent Assay Workflow

The steps below represent a typical workflow for using the PerkinElmerOpal 4-Color and 7-Color Automation IHC Kits on the Leica Biosystems BOND RX.

- 1 Optimize the Opal assay setup on page 13
- 2 Prepare the Materials on page 13
- 3 Set up the BOND RX Protocols on page 18
- 4 Run the Opal 4-Color or 7-Color Automation IHC Kit on page 25

NOTE



PerkinElmer recommends that you run Library slides and Autofluorescence slides with the tissue samples in every run.

Optimize the Opal assay setup

Refer to PerkinElmer's Opal assay development guide.

Prepare the Materials

Table 4. Materials Required

Materials provided by PerkinElmer	Materials provided by Leica Biosystems	Materials provided by User
Antibody Diluent/Block	Stainer	• 1° Antibodies
Opal Polymer HRP Ms+Rb	Leica Biosystems' BOND RX System	• 1xTBS
• 1X Plus Amplification Diluent	Bulk Reagents	• 100% Ethanol
Opal 520 Reagent	BOND Epitope Retrieval Solution 1	Deionized or Milli-Q Water
Opal 540 Reagent	BOND Epitope Retrieval Solution 2	ProLong® Diamond Antifade Mountant
Opal 570 Reagent	BOND Wash Solution 10X	Cover Glass, No. 1.5
Opal 620 Reagent	BOND Dewax Solution	Tissue-Tek® Vertical 24 Slide Rack
Opal 650 Reagent	Consumables	Tissue-Tek® Staining Dish
Opal 690 Reagent	BOND Titration Kit	
Spectral DAPI	BOND Open Containers 30 mL	
• DMSO	BOND Research Detection System 2	
	BOND Universal Covertiles	

Prepare the Reagents

- Reconstitute each dry vial of Opal fluorophore with 75 µL of the DMSO provided. Aliquot the Opal fluorophores if necessary. Opal fluorophores must be stored at 4 °C after reconstituted.
- Allow the reconstituted Opal fluorophore to unfreeze (if taken out of refrigerator), by allowing it to sit at room temperature for 10 min.
 When unfrozen, shake and spin down the vials before diluting the quantity needed for a run at 1:150 with the 1X Plus Amplification Diluent.

NOTE



Ensure antibodies are diluted fresh prior to starting your run.

Testing suggests a good signal balancing when using a dilution of 1:150 (Opal dye: Amplification Diluent). This is a good to place to start; however, some antibodies/targets may require more or less concentrated Opal fluorophore.

- Dilute all antibodies with the Antibody Diluent/Blocking (ARD1001EA) provided, using antibody concentrations obtained from previously conducted titration experiments.
- Dilute the Spectral DAPI (FP1490) at a concentration of 1 drop of DAPI to every 500 µL of TBS.

NOTE



Do not introduce bubbles into the solutions by shaking the containers. To mix reagents, gently invert or vortex the containers several times. If bubbles are present, leave the containers out at room temperature until the bubbles dissipate.

Prepare the BOND RX Instrument

Refer to BOND RX User Manual

Add Opal Detection Buffer Reagent

(Optional: for those who are willing to designate a specific Research Detection System for Opal protocol)

NOTE



This section is made for those who do not already have a **Bond Research Detection System** made; Skip this step if you may want to use your own detection system.

The **Opal Detection Buffer reagent** will be used as the reagent to register with the detection system.



- 1 In the BOND RX software, click the Reagent Setup icon at the top of the System Status Screen.
- 2 Click the Add button. The Add Reagent window opens.
- 3 Type Opal Detection Buffer in the Name text box.
- 4 Type **Buffer** in the **Abbreviated Name** text box.
- 5 Select **Ancillary** in the **Type** drop-down list.
- 6 Leave the **Supplier** text box empty for this reagent.
- 7 Click Save.

Register and Prepare the Opal Detection System on the BOND RX

NOTE



This section is made for those who do not already have a **Bond Research Detection System** made;

Skip this step if you may want to use your own detection system.

- 1 Obtain a new BOND Research Detection System.
- **2** Scan the barcode located on the front of the detection system.
- 3 Enter OPAL Detection Kit into the Name text box.
- 4 Select the first reagent slot on the right, go to the drop down and select **Opal Detection Buffer**.

- 5 Scan a new 30 mL open container and place it into the first position on the detection system.
- 6 Tape the container into place on the detection system and label it as **Opal Detection Buffer**.
- 7 Hit **OK**.

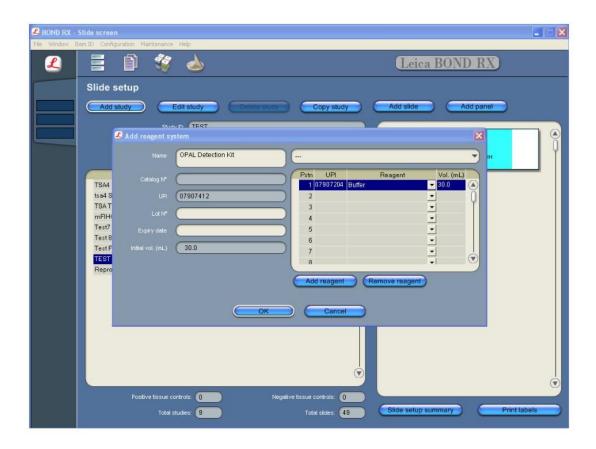


Figure 1. Register Opal Detection System Window

Prepare the Instrument Reagents

NOTE



When preparing to start a run, fill the BOND RX containers with the appropriate reagents according to Table 5. *Open 1-6 can be modified to fit the antibody containers that the user may already have.

Table 5. Reagent Names in BOND RX

Reagent to Add	Container Name	Abbreviated Name	Supplier	Container Size
TBS	Opal Detection Buffer	Buffer	User	30 mL
Antibody Diluent/Block	*PKI Blocking Buffer	PKIBlock	PerkinElmer	30 mL
Antibody 1 with Antibody Diluent	*Open 1	Open 1	User/PerkinElmer	6 mL
Antibody 2 with Antibody Diluent	*Open 2	Open 2	User/PerkinElmer	6 mL
Antibody 3 with Antibody Diluent	*Open 3	Open 3	User/PerkinElmer	6 mL
Antibody 4 with Antibody Diluent	*Open 4	Open 4	User/PerkinElmer	6 mL
Antibody 5 with Antibody Diluent	*Open 5	Open 5	User/PerkinElmer	6 mL
Antibody 6 with Antibody Diluent	*Open 6	Open 6	User/PerkinElmer	6 mL
Opal Polymer HRP Ms+Rb	*Opal Polymer HRP	Opal HRP	PerkinElmer	30 mL
Opal 520 with 1X Plus Amplification Diluent	*Opal 520 Reagent	Opal 520	PerkinElmer	6 mL
Opal 540 with 1X Plus Amplification Diluent	*Opal 540 Reagent	Opal 540	PerkinElmer	6 mL
Opal 570 with 1X Plus Amplification Diluent	*Opal 570 Reagent	Opal 570	PerkinElmer	6 mL
Opal 620 with 1X Plus Amplification Diluent	*Opal 620 Reagent	Opal 620	PerkinElmer	6 mL
Opal 650 with 1X Plus Amplification Diluent	*Opal 650 Reagent	Opal 650	PerkinElmer	6 mL
Opal 690 with 1X Plus Amplification Diluent	*Opal 690 Reagent	Opal 690	PerkinElmer	6 mL
Spectral DAPI with TBS	*Spectral DAPI	DAPI	PerkinElmer	6 mL
TBS	TBS	TBS	User	30 mL

NOTE



Do not introduce bubbles into the solutions by shaking the containers. To mix reagents, gently invert the containers several times. If bubbles are present, leave the containers out at room temperature until the bubbles dissipate.

Set up the BOND RX Protocols

Perform the following procedures to set up the BOND RX:

- Create a Fake Probe Hybridization Protocol on page 18
- Create a Fake Probe Reagent on page 19
- Create a Staining Protocol on page 20
- Create an Autofluorescence Protocol on page 21
- Create a Library Protocol on page 21
- Create a DAPI Protocol on page 22
- Setup a Study on page 23

NOTE



Users with the **BOND RX Version 5.2** software can skip all fake probe steps.

Create a Fake Probe Hybridization Protocol

- 1 In the **Protocol Setup** window, select **Prestaining** under Protocol Group and **ISH Hybridization** under the Protocol Type menu.
- 2 Select the *ISH Hybridization (2Hr) protocol and click Copy.
- 3 Change the Name to OPAL IHC FAKE HYB

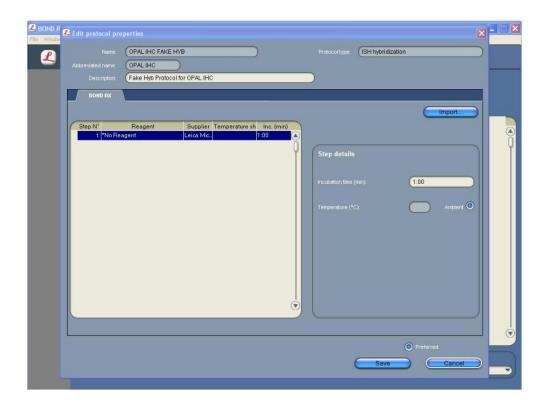


Figure 2. Copy Protocol Window

- Change the Abbreviated Name to **OPAL IHC**.
- Change the Incubation Time to 1 minute. 5
- Change the Temperature to Ambient.
- 7 Click Save.

Create a Fake Probe Reagent

- Click the **Reagent Setup** icon at the top of the window.
- 2 Click **Add** to open the Add Reagent window.
- 3 Type **OPAL IHC** in the Name text box.
- Type **OPAL IHC** in the Abbreviated Name text box.
- 5 Select **Probe** in the Type drop-down list.
- Type **Fake Probe** in the Supplier text box.
- Check **RNA** as Probe Type.

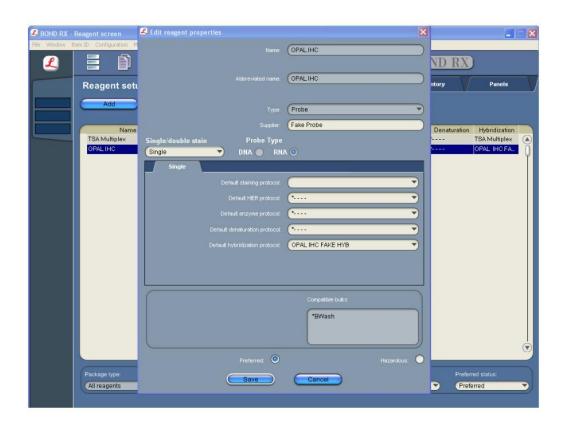


Figure 3. Add Reagent Window

- 8 Select **OPAL IHC FAKE HYB** as the default hybridization protocol.
- 9 Click Save.
- 10 Register a 30 mL Open Container under the new reagent **OPAL IHC** (Fake Probe) and fill the container with TBS.

Create a Staining Protocol

- 1 In the Protocol Setup window, select Protocol Group->Staining.
- 2 Select the **Opal 7-Color** protocol and press **Copy**.
 - If you are using Opal 7-Color Automation Kit, ensure the asterisk in the protocol name and abbreviated name is deleted and go to step 4.
 - If you are using Opal 4-Color Automation Kit, go to step 3.
- 3 For Opal 4-Color Automation Kit: Truncate the protocol for a 3-plex assay by deleting steps related to Ab2-Opal 540, Ab4-Opal 620 and Ab5-Opal 650. Name the new protocol as **Opal 4-Color** and change the abbreviated name to **PKI4c**.
- 4 Select **Opal Detection Kit** (or your preferred detection system) under the preferred detection system menu.
- 5 Replace first Bond Wash Solution step as a reagent step (figure 4 below) and input the reagent linked to your detection system for 0 minutes (Opal Detection Buffer for those who created their detection system with this manual). Alternatively, if you are using your own detection system and reagent, ensure the reagent linked to your detection system has a step within the 7-color or 4-color protocol.
- 6 Click **Show Wash Steps** and compare the protocol to the steps in Appendix A to ensure all steps are present and correct.
- 7 Click Save.



Figure 4. Add Detection System Buffer Step

Create an Autofluorescence Protocol

- 1 In the Protocol setup window, select **Protocol Group -> Staining**.
- 2 Select either the **OPAL 4-Color** or **OPAL 7-Color** protocol created above and click **Copy**.
- 3 Change the Name to OPAL 4-Color AF or OPAL 7-Color AF.
- 4 Change the Abbreviated Name to 4plex AF or 6plex AF.
- 5 Select **OPAL Detection Kit** under the preferred detection system menu.
- 6 Change all 6 Opal fluorophore steps as well as DAPI to the **TBS** container, and leave all other steps the same.
- 7 Click Save.

Create a Library Protocol

- 1 In the Protocol Setup window, select Protocol Group -> Staining.
- 2 Select either the OPAL 4-Color or OPAL 7-Color protocol created above and click Copy.

- 3 Change the Name to **OPAL 4-Color** or **OPAL 7-Color Ab1** Library.
- 4 Change the Abbreviated Name to 3pxAb1Lb or 6pxAb1Lb.
- 5 Select **OPAL Detection Kit** under the preferred detection system menu.
- 6 Select all non-stripping reagent steps that are not involved with cycle of the library antibody, except the initial Buffer step, and change them to the **TBS container**.

Example: For the library protocol of antibody 1: PKI Blocking (step 3), Antibody 1 (step 4), PKI HRP (step 8), and OPAL 520 (step 14) will remain unchanged. All other blocking, antibody, HRP, Opal fluorophore and DAPI steps will be changed to TBS.

- 7 Click Save.
- **8** Repeat for all other antibodies to produce 3 or 6 library slide protocols, depending on if you are using the 4-Color or 7-color protocol.

Create a DAPI Protocol

- 1 In the Protocol Setup window, select Protocol Group -> Staining.
- 2 Select either the OPAL 4-Color or OPAL 7-Color protocol created above and click Copy.
- 3 Change the Name to OPAL 4-Color DAPI or OPAL 7-Color DAPI.
- 4 Change the Abbreviated Name to 3px DAPI or 6px DAPI.
- 5 Select **OPAL Detection Kit** under the preferred detection system menu.
- 6 Select all non-stripping reagent steps, except the initial Buffer step and DAPI, and change them to **TBS**.
- 7 Click Save.

Set Up a Study

To create a study in the BOND RX software:

- 1 Click the **Slide Setup** icon on the BOND RX System Status window.
- 2 Click the **Add Study** button. The Add Study window opens (see figure 5 below).
- 3 Type a name in the Study ID text box.
- 4 Select 150 μL in the Dispense Volume drop-down list.
- 5 Select *Bake and Dewax or *Dewax in the Preparation Protocol drop-down list, depending on whether the slides need to be baked.
- 6 Click OK.



Figure 5. Add Study Window

- 7 Click the **Add Slide** button to assign a protocol to each slide in the study. The Add Slide window opens (see figure 6 below).
- 8 Type the name of the tissue in the **Comments** text box.
- 9 Select ISH as the Process on the Research tab.
- 10 Select the marker **OPAL IHC (Fake Probe)**, if using Version 4 software.
- 11 In the **Preparation Protocol** drop-down list, verify the correct

- protocol, either *Bake and Dewax or *Dewax, is selected.
- 12 In the Staining Protocol drop-down list, select OPAL 4-Color or OPAL 7-Color, or other appropriate protocol.
- 13 Select the HIER Protocol based on the initial antibody in the 6plex. For an antibody that typically receives a pH6 epitope retrieval step, select *HIER 20 min with ER1. For an antibody that typically receives a pH9 epitope retrieval step, select *HIER 20 min with ER2. (For multiplex staining on version 4.0, it's recommended to start with an antibody that needs ER2 for epitope retrieval.)
- 14 Select OPAL IHC FAKE HYB as the Hybridization protocol.
- 15 Leave all other text boxes empty.
- 16 Click the Add Slide button to add the slide to the study.

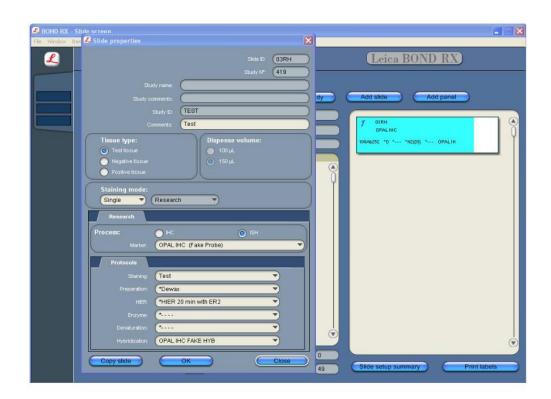


Figure 6. Add Slide Window

- 17 Repeat steps 7 to 16 to add each of the slides to the protocol.
- **18** When all of the slides are added, click the **Close** button to close the Add Slide window.
- 19 Repeat the procedure above to add autofluorescent or library slides.
- **20** Click the **Print Labels** button at the bottom of the Slide Setup window to print a barcode label for each slide.

Run the Opal 4-Color or 7-Color Automation IHC Kit

- Materials Required on page 25
- Load the Reagents on page 25
- Start the Run on page 26
- Complete the Run on page 26

Materials Required

The following materials are required to run the assay:

- TBS
- Water (DI, MQ)
- Cover Glass, No. 1.5
- ProLong[®] Diamond Antifade Mountant
- Slide Rack
- Staining Dish

Load the Reagents

- 1 Label each BOND RX container with the corresponding reagent from Table 5, using a water-resistant marker.
- 2 Use the barcode scanner to scan the barcode on the **front** of each container. A pop-up menu will appear.

NOTE



Do NOT scan the barcode on the top of the container.

- 3 Choose the appropriate Reagent Name, and enter the lot number and expiration date in the corresponding text boxes. Click OK.
- **4** Load the containers onto the reagent tray and slide the tray into the BOND RX.

Start the Run

1 After attaching the barcode labels to the slides, add the slides to the slide tray with the labels facing up.

NOTE



Autofluorescent, library, and 4-Color or 7-Color protocols may be able to run on the same tray, assuming that the only differences are the reagents that are dispensed during each step, and each step across the protocols is the same amount of time.

- 2 Add a covertile on top of each slide. The rectangular-shaped neck of the covertile should fit into the groove of the slide tray.
- 3 Place the tray in the BOND RX and press the button to load the tray onto the machine.
- 4 After the slides have been scanned, go to the **System Status Screen** and select the triangular (**PLAY**) button on the screen under the tray to start the run. Alternatively, right-click on the scanned label images and select **Delayed Start** to start the run at a future time.

NOTE



Typical time for a 6plex run will take 11-13 hours, depending on the amount of slides being run. It is recommended that the delayed start is used if slides may be left on the BOND RX for more than 10 minutes beyond run completion.

Before leaving the instrument unattended, verify that the instrument is running properly, and that an end time has been scheduled.

Complete the Run

- 1 After the run is complete, press the button on the instrument to unload the slides.
- 2 Place the Slide Rack into the staining dish, and fill the dish with DI water.
- **3** Remove the covertiles and place in a separate container to clean.
- 4 Place the slides into the Slide Rack inside the staining dish.

NOTE



Do not leave slides on the BOND RX after the run is completed. Promptly remove and coverslip for best staining results.

- **5** Remove the slides one at a time from the Slide Rack and wipe off excess water on the sides of the tissue and on the back of the slide.
- 6 Mount by adding 1-2 DROPS of ProLong DiamondAntifade mounting medium to each slide and carefully placing a No. 1.5 coverglass over the section. Avoid trapping air bubbles.
- **7** Air dry slides for at least 20 min.
- **8** Examine the slide results on the Vectra Automated Quantitative Pathology Workstation.

Appendix A: Opal 7-Color Protocol

	Appendix A.	<u> </u>		10001
Step No.	Reagent	Step Type	Incubation Time	Temperature
1	Opal Detection Buffer	Reagent	0 min	Ambient
2	Bond Wash Solution	Wash	0 min	Ambient
3	PKI Blocking Buffer	Reagent	5 min	Ambient
4	Open 1	Reagent	30 min	Ambient
5	Bond Wash Solution	Wash	0 min	Ambient
6	Bond Wash Solution	Wash	1 min	Ambient
7	Bond Wash Solution	Wash	0 min	Ambient
8	Opal Polymer HRP	Reagent	10 min	Ambient
9	Bond Wash Solution	Wash	0 min	Ambient
10	Bond Wash Solution	Wash	1 min	Ambient
11	Bond Wash Solution	Wash	0 min	Ambient
12	Bond Wash Solution	Wash	0 min	Ambient
13	Bond Wash Solution	Wash	0 min	Ambient
14	Opal 520 Reagent	Reagent	10 min	Ambient
15	Bond Wash Solution	Wash	0 min	Ambient
16	Bond Wash Solution	Wash	1 min	Ambient
17	Bond Wash Solution	Wash	0 min	Ambient
18	Bond Wash Solution	Wash	0 min	Ambient
19	Bond ER1 Solution	Reagent	0 min	Ambient
20	Bond ER1 Solution	Reagent	0 min	95 °C
21	Bond ER1 Solution	Reagent	20 min	95 °C
22	Bond ER1 Solution	Reagent	0 min	Ambient
23	Bond Wash Solution	Wash	0 min	Ambient
24	Bond Wash Solution	Wash	1 min	Ambient
25	Bond Wash Solution	Wash	0 min	Ambient
26	PKI Blocking Buffer	Reagent	5 min	Ambient
27	Open 2	Reagent	30 min	Ambient
28	Bond Wash Solution	Wash	0 min	Ambient
29	Bond Wash Solution	Wash	1 min	Ambient
30	Bond Wash Solution	Wash	0 min	Ambient
31	Opal Polymer HRP	Reagent	10 min	Ambient
32	Bond Wash Solution	Wash	0 min	Ambient
33	Bond Wash Solution	Wash	1 min	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature
34	Bond Wash Solution	Wash	0 min	Ambient
35	Bond Wash Solution	Wash	0 min	Ambient
36	Bond Wash Solution	Wash	0 min	Ambient
37	Opal 540 Reagent	Reagent	10 min	Ambient
38	Bond Wash Solution	Wash	0 min	Ambient
39	Bond Wash Solution	Wash	1 min	Ambient
40	Bond Wash Solution	Wash	0 min	Ambient
41	Bond Wash Solution	Wash	0 min	Ambient
42	Bond ER1 Solution	Reagent	0 min	Ambient
43	Bond ER1 Solution	Reagent	0 min	95 °C
44	Bond ER1 Solution	Reagent	20 min	95 °C
45	Bond ER1 Solution	Reagent	0 min	Ambient
46	Bond Wash Solution	Wash	0 min	Ambient
47	Bond Wash Solution	Wash	1 min	Ambient
48	Bond Wash Solution	Wash	0 min	Ambient
49	PKI Blocking buffer	Reagent	5 min	Ambient
50	Open 3	Reagent	30 min	Ambient
51	Bond Wash Solution	Wash	0 min	Ambient
52	Bond Wash Solution	Wash	1 min	Ambient
53	Bond Wash Solution	Wash	0 min	Ambient
54	Opal Polymer HRP	Reagent	10 min	Ambient
55	Bond Wash Solution	Wash	0 min	Ambient
56	Bond Wash Solution	Wash	1 min	Ambient
57	Bond Wash Solution	Wash	0 min	Ambient
58	Bond Wash Solution	Wash	0 min	Ambient
59	Bond Wash Solution	Wash	0 min	Ambient
60	Opal 570 Reagent	Reagent	10 min	Ambient
61	Bond Wash Solution	Wash	0 min	Ambient
62	Bond Wash Solution	Wash	1 min	Ambient
63	Bond Wash Solution	Wash	0 min	Ambient
64	Bond Wash Solution	Wash	0 min	Ambient
65	Bond ER1 Solution	Reagent	0 min	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature
66	Bond ER1 Solution	Reagent	0 min	95 °C
67	Bond ER1 Solution	Reagent	20 min	95 °C
68	Bond ER1 Solution	Reagent	0 min	Ambient
69	Bond Wash Solution	Wash	0 min	Ambient
70	Bond Wash Solution	Wash	1 min	Ambient
71	Bond Wash Solution	Wash	0 min	Ambient
72	PKI Blocking Buffer	Reagent	5 min	Ambient
73	Open 4	Reagent	30 min	Ambient
74	Bond Wash Solution	Wash	0 min	Ambient
75	Bond Wash Solution	Wash	1 min	Ambient
76	Bond Wash Solution	Wash	0 min	Ambient
77	Opal Polymer HRP	Reagent	10 min	Ambient
78	Bond Wash Solution	Wash	0 min	Ambient
79	Bond Wash Solution	Wash	1 min	Ambient
80	Bond Wash Solution	Wash	0 min	Ambient
81	Bond Wash Solution	Wash	0 min	Ambient
82	Bond Wash Solution	Wash	0 min	Ambient
83	Opal 620 Reagent	Reagent	10 min	Ambient
84	Bond Wash Solution	Wash	0 min	Ambient
85	Bond Wash Solution	Wash	1 min	Ambient
86	Bond Wash Solution	Wash	0 min	Ambient
87	Bond Wash Solution	Wash	0 min	Ambient
88	Bond ER1 Solution	Reagent	0 min	Ambient
89	Bond ER1 Solution	Reagent	0 min	95 °C
90	Bond ER1 Solution	Reagent	20 min	95 °C
91	Bond ER1 Solution	Reagent	0 min	Ambient
92	Bond Wash Solution	Wash	0 min	Ambient
93	Bond Wash Solution	Wash	1 min	Ambient
94	Bond Wash Solution	Wash	0 min	Ambient
95	PKI Blocking Buffer	Reagent	5 min	Ambient
96	Open 5	Reagent	30 min	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature
97	Bond Wash Solution	Wash	0 min	Ambient
98	Bond Wash Solution	Wash	1 min	Ambient
99	Bond Wash Solution	Wash	0 min	Ambient
100	Opal Polymer HRP	Reagent	10 min	Ambient
101	Bond Wash Solution	Wash	0 min	Ambient
102	Bond Wash Solution	Wash	1 min	Ambient
103	Bond Wash Solution	Wash	0 min	Ambient
104	Bond Wash Solution	Wash	0 min	Ambient
105	Bond Wash Solution	Wash	0 min	Ambient
106	Opal 650 Reagent	Reagent	10 min	Ambient
107	Bond Wash Solution	Wash	0 min	Ambient
108	Bond Wash Solution	Wash	1 min	Ambient
109	Bond Wash Solution	Wash	0 min	Ambient
110	Bond Wash Solution	Wash	0 min	Ambient
111	Bond ER1 Solution	Reagent	0 min	Ambient
112	Bond ER1 Solution	Reagent	0 min	95 °C
113	Bond ER1 Solution	Reagent	20 min	95 °C
114	Bond ER1 Solution	Reagent	0 min	Ambient
115	Bond Wash Solution	Wash	0 min	Ambient
116	Bond Wash Solution	Wash	1 min	Ambient
117	Bond Wash Solution	Wash	0 min	Ambient
118	PKI Blocking Buffer	Reagent	5 min	Ambient
119	Open 6	Reagent	30 min	Ambient
120	Bond Wash Solution	Wash	0 min	Ambient
121	Bond Wash Solution	Wash	1 min	Ambient
122	Bond Wash Solution	Wash	0 min	Ambient
123	Opal Polymer HRP	Reagent	10 min	Ambient
124	Bond Wash Solution	Wash	0 min	Ambient
125	Bond Wash Solution	Wash	1 min	Ambient
126	Bond Wash Solution	Wash	0 min	Ambient
127	Bond Wash Solution	Wash	0 min	Ambient

Step No.	Reagent	Step Type	Incubation Time	Temperature
128	Bond Wash Solution	Wash	0 min	Ambient
129	Opal 690 Reagent	Reagent	10 min	Ambient
130	Bond Wash Solution	Wash	0 min	Ambient
131	Bond Wash Solution	Wash	1 min	Ambient
132	Bond Wash Solution	Wash	0 min	Ambient
133	Bond Wash Solution	Wash	0 min	Ambient
134	Bond ER1 Solution	Reagent	0 min	Ambient
135	Bond ER1 Solution	Reagent	0 min	95 °C
136	Bond ER1 Solution	Reagent	20 min	95 °C
137	Bond ER1 Solution	Reagent	0 min	Ambient
138	Bond Wash Solution	Wash	0 min	Ambient
139	Bond Wash Solution	Wash	1 min	Ambient
140	Bond Wash Solution	Wash	0 min	Ambient
141	Spectral DAPI	Reagent	5 min	Ambient
142	Bond Wash Solution	Wash	0 min	Ambient
143	Bond Wash Solution	Wash	1 min	Ambient
144	Bond Wash Solution	Wash	0 min	Ambient

Appendix B: Safety

The following safety information about the Opal 4-Color and OPAL 7-Color Automation IHC Kits is included in this documentation. Read and review all safety information before operating the Instrument or using the reagents.

- Required Training on page 33
- Chemical and Biological Safety on page 33

Required Training

Ensure that all personnel involved with the operation of the instrument have:

- Received instruction in general safety practices for laboratories.
- Received instruction in specific safety practices for the instrument.
- Read and understood all related MSDSs.

WARNING



Use this product only in the manner described in this manual. If the equipment is used in a manner not specified by the manufacturer, the protection provided by the equipment may be impaired.

Chemical and Biological Safety

WARNING



Biohazard Risks. In some applications, chemicals or samples used with the Opal 4-Color and 7-Color Automation IHC Kit are potentially hazardous and cancause illness.

- Read and understand the material safety data sheet (MSDS) provided by the chemical manufacturer before you store, handle, or work with any chemical or hazardous material.
- Minimize contact with and inhalation of chemicals and chemical wastes.
- Wear appropriate personal protective equipment when handling chemicals or samples (e.g., safety glasses, gloves, or clothing). For additional safety guidelines consult the MSDS.
- Handle all samples using good laboratory practices to prevent biohazards.

Chemical and Biological Safety (Continued)

- Do not leave chemical containers open. Use only with adequate ventilation, including a fume hood, if necessary.
- Check regularly for chemical leaks or spills. If a leak or spill occurs, follow the chemical manufacturer's cleanup procedures as recommended on the MSDS.
- Dispose of chemical or infectious waste in accordance with good laboratory practices and local, state/provincial, or national environmental and health regulations.
- After emptying waste containers, seal the waste containers appropriately.
- Comply with all local, state/provincial, or national laws and regulations related to chemical and waste storage, handling, and disposal.

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